

Inventors:

Attorney's Docket No.: GSF97-03

RELATED APPLICATIONS

This is a continuation application of PCT/EP96/01001
5 filed March 8, 1996, which claims priority to Danish patent
application DK 0243/95 filed March 9, 1995. The contents
of PCT/EP96/01001 and DK 0243/95 are incorporated herein by
reference in their entirety.

BACKGROUND OF THE INVENTION

- 10 The introduction of therapeutic genes into cells for
the treatment of diseases as diverse as those resulting
from genetic defects, cancer and viral infections is the
major aim of gene therapy. Cancer and diseases such as
AIDS resulting from infection with human immunodeficiency
15 virus (HIV) are particularly difficult to treat even though
a number of clinical protocols are presently underway that
use gene therapeutical approaches. The amphipathic peptide
melittin, the major component of bee venom, has been shown
to have selective anti-cancer (Sharma, S.V., *Oncogene*,
20 7:193-201 (1992); Sharma, S.V., *Oncogene*, 8:939-947 (1993))
and anti-HIV activity (Wachinger et al., *FEBS Lett.*,
309:235-241 (1992)); U.S. Patent No. 4,822,608 of Benton et

U.S. Patent No. 4,822,608 issued to Benton et al. on April 18, 1989 and entitled "METHODS AND COMPOSITIONS FOR THE TREATMENT OF MAMMALIAN INFECTIONS EMPLOYING MEDICAMENTS COMPRISING HYMENOPTERA VENOM OR PROTEINACEOUS OR POLYPEPTIDE COMPONENTS THEREOF" teaches that secondary agents derived from nature such as hymenoptera venom or proteinaceous or polypeptide components thereof has a potentiating effect on antibacterial agents. This reference further suggests that such compositions may also have increased anti-viral, carcinostatic and anti-carcinogenic effects on various maladies. More particularly, the reference to Benton et al. discloses the use of melittin which is the main component of honey bee toxin, in combination with assorted antibiotic agent as having antibacterial activity against predetermined infections. Further this reference teaches that a synergistic benefit may be achieved by the combination of the melittin and assorted antibiotics in various therapeutically effective amounts.

In these studies purified melittin peptide was given to cells in culture which, though useful for experimental purposes, is not relevant for therapy. Even in vivo

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administration of purified melittin protein (for example i.v.) is probably not advisable because of the relatively high concentrations and repeated doses that would be required to maintain therapeutic levels. Further, since
5 this kind of generalized delivery would result in the amphipathic peptide reaching not only target cells but also other cells, thereby potentially resulting in nondesirable side effects, it would be advantageous to be able to target the delivery of melittin or other antimicrobial peptides,
10 in particular the melittin peptide and the peptides mentioned below.

A second class of therapeutic genes of interest are the cecropins isolated from the pupae of giant silk moths (Bowman, H.G., *Ann. Rev. Immunol.*, 13:1-51 (1995), Hultmark
15 et al., *Eur. J. Biochem.*, 106:7-16 (1980)). There are three principal cecropins, A, B and D with a similar structure to melittin (Hultmark et al., *Eur. J. Biochem.*, 127:207-217 (1982)). Cecropins A and B show specific antibacterial activity without any apparent ill effects for
20 mammalian cells (Steiner H. et al., *Nature*, 292:246-248 (1991)). Recently Moore and coworkers have shown that the cecropin B, P and Shiva-1 antibacterial peptides show anticancer activity against a variety of tumour cell lines (Moore, A.J. et al., *Peptide Research*, 7:265-269 (1994)).

25 Cecropins were first isolated from the hemolymph of the giant silk moth, *Hyalophora cecropia*, following induction by live non-pathogenic bacteria. The principal insect cecropins (A, B and D) are 35 to 37 residues long, devoid of cysteine and have a strongly basic N-terminus
30 linked to a neutral C-terminus by a flexible glycine-proline link. The overall structure deduced by NMR for cecropin A is two nearly perfect amphipathic segments joined by a Gly-Pro hinge. A cecropin-like 31-residue

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peptide (cecropin P₁), isolated from the small intestine of a pig (Lee et al., *Proc. Natl. Acad. Sci. USA*, 86:9159-9162 (1989)), suggests that the cecropins may be widespread throughout the animal kingdom. The mechanism of action of the cecropins is thought to involve channel formation in membranes and subsequent lysis.

SB-37 (a close cecropin B analogue) and Shiva-1 (a cecropin B analogue that shares about 40% sequence homology and maintains the same charge distribution and hydrophobicity as the peptide) have been shown to lyse several mammalian leukemia and lymphoma cell lines *in vitro*. The publication of Moore, A.J. et al., *Peptide Research*, 7:265-269 (1994) is incorporated herein by reference for complete disclosure. Similar antitumour effects have been demonstrated for the magainins, a related group of antimicrobial peptides (Cruciani, R.A. et al., *Proc. Natl. Acad. Sci. USA*, 88:3792-3796 (1991); Ohaski, Y. et al., *Cancer Research*, 52:3534-3538, (1992)).

The use of retroviral vectors (RV) for gene therapy has received much attention and currently is the method of choice for the transferral of therapeutic genes in a variety of approved protocols both in the USA and in Europe (Kotani, H. et al., *Human Gene Therapy*, 5:19-28 (1994)). However most of these protocols require that the infection of target cells with the RV carrying the therapeutic gene occurs *in vitro*, and successfully infected cells are then returned to the affected individual (Rosenberg, S.A. et al., *Human Gene Therapy*, 3:75-90 (1992); for a review see Anderson, W.F., *Science*, 256:808-813 (1992)). Such *ex vivo* gene therapy protocols are ideal for correction of medical conditions in which the target cell population can be easily isolated (e.g. lymphocytes). Additionally the *ex vivo* infection of target cells allows the administration of

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Unfortunately, only a fraction of the possible applications for gene therapy involve target cells that can be easily isolated, cultured and then reintroduced.

15 This kind of *in vivo* approach, of course, introduces a
variety of new problems. First of all, and above all,
safety considerations have to be addressed. Virus will be
produced, possibly from an implantation of virus producing
cells, and there will be no opportunity to precheck the
produced virus. It is important to be aware of the finite
20 risk involved in the use of such systems, as well as trying
to produce new systems that minimize this risk.

The essentially random integration of the proviral form of the retroviral genome into the genome of the infected cell led to the identification of a number of cellular proto-oncogenes by virtue of their insertional activation (Varmus, H., *Science*, 240:1427-1435 (1988)). The possibility that a similar mechanism may cause cancers in patients treated with RVs carrying therapeutic genes intended to treat other pre-existent medical conditions, has posed a recurring ethical problem. Most researchers would agree that the probability of a replication defective RV, such as all those currently used, integrating into or near a cellular gene involved in controlling cell

proliferation is vanishingly small. However, it is generally also assumed that the explosive expansion of a population of replication competent retrovirus from a single infection event, will eventually provide enough
5 integration events to make such a phenotypic integration a very real possibility.

Retroviral vector systems are optimized to minimize the chance of replication competent virus being present. However, it has been well documented that recombination
10 events between components of the RV system can lead to the generation of potentially pathogenic replication competent virus and a number of generations of vector systems have been constructed to minimize this risk of recombination (reviewed in Salmons, B. and Günzburg, W.H., *Human Gene*
15 *Therapy*, 4:129-141 (1993)). However little is known about the finite probability of these events. Since it will never be possible to reduce the risk associated with this or other viral vector systems to zero, an informed risk-benefit decision will always have to be taken. Thus it
20 becomes very important to empirically determine the chance of (1) insertional disruption or activation of single genes by retrovirus integration and (2) the risk of generation of replication competent virus by recombination in current generations of packaging cell lines. A detailed
25 examination of the mechanism by which these events occur will also allow the construction of new types of systems designed to limit these events.

A further consideration for practical *in vivo* gene therapy, both from safety considerations as well as from an
30 efficiency and from a purely practical point of view, is the targeting of RVs. It is clear that therapeutic genes carried by vectors should not be indiscriminately expressed in all tissues and cells, but rather only in the requisite

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target cell. This is especially important if the genes to be transferred are toxin genes aimed at ablating specific tumour cells. Ablation of other, nontarget cells would obviously be very undesirable. Targeting of the expression of carried therapeutic genes can be achieved by a variety of means.

Retroviral vector systems consist of two components (Figure 1):

1) the retroviral vector itself is a modified retrovirus (vector plasmid) in which the genes encoding for the viral proteins have been replaced by therapeutic genes optionally including marker genes to be transferred to the target cell. Since the replacement of the genes encoding for the viral proteins effectively cripples the virus it must be rescued by the second component in the system which provides the missing viral proteins to the modified retrovirus.

The second component is:

2) a cell line that produces large quantities of the viral proteins, however lacks the ability to produce replication competent virus. This cell line is known as the packaging cell line and consists of a cell line transfected with a second plasmid carrying the genes enabling the modified retroviral vector to be packaged. This plasmid directs the synthesis of the necessary viral proteins required for virion production.

To generate the packaged vector, the vector plasmid is transfected into the packaging cell line. Under these conditions the modified retroviral genome including the

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inserted therapeutic and optional marker genes is transcribed from the vector plasmid and packaged into the modified retroviral particles (recombinant viral particles). A cell infected with such a recombinant viral particle cannot produce new vector virus since no viral proteins are present in these cells. However the vector carrying the therapeutic and marker genes is present and these can now be expressed in the infected cell.

SUMMARY OF THE INVENTION

10 It is an object of the present invention to provide a novel therapeutic agent with antitumour, antiviral, antibacterial and/or antifungal activities.

It is a further object of the present invention to provide a novel therapeutic agent with high selectivity for
15 selected target cells and reduced nondesirable side effects.

To achieve the foregoing and other objects, the present invention provides a recombinant vector for introducing into an eucaryotic cell DNA, the vector
20 comprising, in operable linkage, a) the DNA of or corresponding to at least a portion of a vector, which portion is capable of infecting and directing the expression in the target cells; and b) one or more coding sequences wherein at least one sequence encodes for at
25 least one naturally occurring therapeutic antimicrobial peptide or a derivative thereof for the treatment of at least one disease, selected from mammalian tumours, viral, bacterial and fungal infections. Said sequence is preferably replication-defective.

30 Said sequence encoding a naturally occurring therapeutic antimicrobial peptide or derivative thereof encodes for the amino acid sequences of all, part, an

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analogue, homologue, recombinant or combination thereof of such antimicrobial peptide.

Said sequences comprise preferably also non-coding sequences.

5 The antimicrobial peptides or derivatives thereof include but are not limited to those encoding melittin, the various cecropins and magainins. Further included are the apidaecin and defensin peptides or derivatives thereof. These genes may be expressed in their preproform or
10 alternatively in a genetically engineered preform or in another form which renders a biological active peptide or a derivative thereof.

Said sequence is preferably a recombinant molecule coding for the amino acid sequences of all, part, and
15 analogue, homologue, derivative, recombinant or combination thereof of the melittin, cecropin, magainin, apideacin and defensin genes.

As discussed in detail in the prior art reference to Benton et al., melittin, the main component in honey bee
20 toxin is a polypeptide which includes substantially 26 amino acid residues. These amino acid residues include, Gly-Ile-Gly-Ala-Val-Leu-Lys-Val-Leu-Thr-Thr-Gly-Leu-Pro-Ala-Leu-Ile-Ser-Trp-Ile-Lys-Arg-Lys-Arg-Gln-Gln (SEQ ID NO: 1). Moreover, melittin analogues wherein at least the last
25 six (C-terminal) amino acids is altered and replaced by six glycine residues appear to have a therapeutic benefit similar to melittin, these amino acid analogues having a structure of Gly-Ile-Gly-Ala-Val-Leu-Lys-Val-Leu-Thr-Thr-Gly-Leu-Pro-Ala-Leu-Ile-Ser-Trp-Ile-Gly-Gly-Gly-Gly-Gly
30 (SEQ ID NO: 2). The use of melittin for treatment of HIV infections is disclosed in WO patent application 91/08753 of Erfle et al., which is incorporated herein by reference for complete disclosure. According to a preferred

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embodiment of the invention said structural analogue of melittin is Gly-Ile-Gly-Ala-Val-Leu-Lys-Val-Leu-Thr-Thr-Gly-Leu-Pro-Ala-Leu-Ile-Ser-Trp-Ile-Gly-Gly-Gly-Gly-Gly (SEQ ID NO: 2) or, according to a further preferred
5 embodiment, Amfi 1 or 2 and peptides of GP 41 that are melittin-like.

According to a further preferred embodiment of the invention, the sequence encoding a therapeutic
10 antimicrobial peptide is coding for a hymenoptera venom, at least one active protein component of a hymenoptera venom, at least one polypeptide component of a hymenoptera venom, and mixtures thereof.

The hymenoptera gene is preferably selected from the group consisting of genes coding for honey bee venom,
15 bumble bee venom, yellow jacket venom, bald-faced hornet venom, active protein components of said venom, active protein components of said venom, and mixtures thereof.

Furthermore, the structural analogues of melittin include an amphophilic helix with or without signal peptide
20 and activation domains.

In a preferred embodiment, the recombinant vector is selected from viral and plasmid vectors. Examples for viral vectors are RNA and DNA virus vectors. A particularly preferred RNA virus vector is a retrovirus
25 vector, more particularly a procon vector. Examples for DNA virus vectors are adenoviruses, adenovirus associated viruses and herpes viruses derived vectors. The plasmid vectors include all eucaryotic expression vectors.

In a preferred embodiment of the invention, the
30 recombinant vector is a retroviral vector.

The retroviral genome consists of an RNA molecule with the structure R-U5-gag-pol-env-U3-R (Figure 1). During the process of reverse transcription, the U5 region is

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5 duplicated and placed at the right hand end of the
generated DNA molecule, whilst the U3 region is duplicated
and placed at the left hand end of the generated DNA
molecule (Figure 1). The resulting structure U3-R-U5 is
called LTR (Long Terminal Repeat) and is thus identical and
repeated at both ends of the DNA structure or provirus.
The U3 region at the left hand end of the provirus harbours
the promoter. This promoter drives the synthesis of an RNA
transcript initiating at the boundary between the left hand
10 U3 and R regions and terminating at the boundary between
the right hand R and U5 region (Figure 1). This RNA is
packaged into retroviral particles and transported into the
target cell to be infected. In the target cell the RNA
genome is again reverse transcribed as described above.

15 According to another embodiment of the invention a
promoter conversion vector (procon vector) can be
constructed in which the righthand U3 region is altered
(Figure 3), but the normal lefthand U3 structure is
maintained (Figure 3); the vector can be normally
20 transcribed into RNA utilizing the normal retroviral
promoter located within the left hand U3 region (Figure 3).
However the generated RNA will only contain the altered
righthand U3 structure. In the infected target cell, after
reverse transcription, this altered U3 structure will be
25 placed at both ends of the retroviral structure (Figure 3).

30 If the altered region carries a polylinker instead of
the U3 region then any promoter, including those directing
tissue specific expression such as the WAP promoter can be
easily inserted. This promoter will then be utilized
exclusively in the target cell for expression of linked
genes carried by the retroviral vector. Alternatively or
additionally DNA segments homologous to one or more

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cellular sequences can be inserted into the polylinker for the purpose of gene targeting.

In the packaging cell line the expression of the retroviral vector is thus regulated by the normal
5 unselective retroviral promoter (Figure 3). However as soon as the vector enters the target cell promoter conversion occurs, and the therapeutic genes are expressed from a tissue specific promoter of choice introduced into the polylinker (Figure 3). Not only can virtually any
10 tissue specific promoter be included in the system, providing for the selective targeting of a wide variety of different cell types, but additionally, following the conversion event, the structure and properties of the retroviral vector no longer resembles that of a virus.
15 This, of course, has extremely important consequences from a safety point of view, since ordinary or state of the art retroviral vectors readily undergo genetic recombination with the packaging vector to produce potentially pathogenic viruses. Promoter conversion (Procon) vectors do not
20 resemble retroviruses because they no longer carry U3 retroviral promoters after conversion thus reducing the possibility of genetic recombination inserted into the polylinker for the purposes of gene targeting.

For a complete disclosure of the procon vectors, the
25 content of the Danish application DK 1017/94, filed on September 2, 1994 is completely included within the present application or incorporated herein by reference.

Thus in a further preferred embodiment of the invention a retroviral vector undergoing promoter
30 conversion (procon vector) is provided comprising a 5'LTR region of the structure U3-R-U5; one or more coding sequences wherein at least one of said coding sequences encodes a naturally occurring antimicrobial peptide or a

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derivative thereof (part, an analogue, homologue, recombinants or a combination thereof of such antimicrobial gene) for the treatment of at least one disease selected from mammalian tumours, viral infections, bacterial
5 infections and fungal infections; and a 3'LTR region comprising a completely or partially deleted U3 region wherein said deleted U3 region is replaced by a polylinker sequence, followed by the R and U5 region.

And in a further preferred embodiment a retroviral
10 vector is provided wherein said retrovirus vector includes, in operable linkage, a 5'LTR region and a 3'LTR region, said 5'LTR region comprising the structure U3-R-U5 and said 3'LTR region comprising a completely or partially deleted U3 region wherein said deleted U3 region is replaced by one
15 or more of said coding sequences wherein at least one sequence encodes for at least one naturally occurring therapeutic antimicrobial peptide or a derivative thereof for the treatment of at least one disease, selected from mammalian tumours, viral, bacterial and fungal infections
20 expressed from either the viral or a heterologous promoter, followed by the R and U5 region.

According to the invention the term "polylinker" is used for a short stretch of artificially synthesized DNA which carries a number of unique restriction sites allowing
25 the easy insertion of any promoter or DNA segment. The term "heterologous" is used for any combination of DNA sequences that is not normally found intimately associated in nature. According to the invention, the heterologous DNA fragment can encode for example a peptide such as a
30 marker peptide (e.g., β -galactosidase, neomycin, alcohol dehydrogenase, puromycin, hypoxanthine phosphoribosyl transferase (HPRT), hygromycin, and secreted alkaline phosphatase), a therapeutic peptide (e.g., herpes simplex

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With reference to the procon vectors, said polylinker sequence carries at least one unique restriction site and contains preferably at least one insertion of a heterologous DNA fragment. Said heterologous DNA fragment

Gene expression is regulated by promoters. In the
20 absence of promoter function a gene will not be expressed.
The normal MLV retroviral promoter is fairly unselective in
that it is active in most cell types. However a number of
promoters exist that show activity only in very specific
cell types. Such tissue-specific promoters will be the
25 ideal candidates for the regulation of gene expression in
retroviral vectors, limiting expression of the therapeutic
genes to specific target cells.

The target cell specific regulatory elements and promoters are preferably, but not limited to one or more elements of the group consisting of, Whey Acidic Protein (WAP), Mouse Mammary Tumour Virus (MMTV), β -lactoglobulin and casein specific regulatory elements and promoters, which may be used to target human mammary tumours, pancreas

[illegible]

25 The antimicrobial genes of the present invention will
be placed under the transcriptional control of for instance
the HIV promoter or a minimal promoter placed under the
regulation of the HIV tat responsive element (TAR) to
target HIV infected cells. Targeting will be achieved
30 because the HIV promoter is dependent upon the presence of
Tat, and HIV encoded autoregulatory protein (Haseltine, W.A., FASEB J., 5:2349-2360 (1991)). Thus only cells
infected with HIV and therefore expressing Tat will be able

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to produce the amphipathic peptide introduced in the Procon vector (Figure 2). Alternatively, the amphipathic peptide could be expressed from T cell specific promoters such as that from the CD4 or T cell receptor gene. In order to
5 target tumour cells, promoters from genes known to be overexpressed in these cells (for example c-myc, c-fos) may be used.

The antimicrobial genes of the present invention may be placed also under the transcriptional control of other
10 promoters known in the art. Examples for such promoters are of the group of SV40, cytomegalovirus, Rous sarcoma virus, β -actin, HIV-LTR, MMTV-LTR, B or T cell specific promoters, tumour specific promoters and HIV.

The retroviral vector is in one embodiment of the
15 invention a BAG vector (Price, J. et al., *Proc. Natl. Acad. Sci. USA*, 87:156-160 (1987)), but includes also other retroviral vectors.

According to a preferred embodiment of the invention at least one retroviral sequence encoding for a retroviral
20 protein involved in integration of retroviruses is altered or at least partially deleted. Said heterologous DNA fragment is preferably homologous to one or more cellular sequences. The regulatory elements and promoters are preferably regulatable by transacting molecules.

25 In a further embodiment of the invention a retroviral vector system is provided comprising a retroviral vector as described above as a first component and a packaging cell line harbouring at least one retroviral or recombinant retroviral construct coding for proteins required for said
30 retroviral vector to be packaged.

The packaging cell line harbours retroviral or recombinant retroviral constructs coding for those retroviral proteins which are not encoded in said

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retroviral vector. The packaging cell line is preferably selected from an element of the group consisting of ψ 2, ψ -Crip, ψ -AM, GP+E-86, PA317 and GP+envAM-12.

After replicating the retroviral vector of the invention as described above in a retroviral vector system as described above, a retroviral provirus is provided wherein said polylinker and any sequences inserted in said polylinker in the 3'LTR become duplicated during the process of reverse transcription in the infected target cell and appear in the 5'LTR as well as in the 3'LTR of the resulting provirus.

The retroviral vector of the invention refers to a DNA sequence retroviral vector on the DNA sequence level.

The invention includes, however, also a retroviral provirus and mRNA of a retroviral provirus according to the invention and any RNA resulting from a retroviral vector according to the invention and cDNAs thereof.

A further embodiment of the invention provides non-therapeutical or therapeutical method for introducing homologous and/or heterologous nucleotide sequences into human or animal cells *in vitro* and *in vivo* comprising transfecting a packaging cell line of a retroviral vector system according to the invention with a retroviral vector according to the invention and infecting a target cell population with recombinant retroviruses produced by the packaging cell line. The nucleotide sequences are selected from one or more elements of the group consisting of genes or parts of genes encoding for therapeutic antimicrobial peptides, regulatory sequences and promoters.

The retroviral vector, the retroviral vector system and the retroviral provirus as well as RNA thereof is used for producing a pharmaceutical composition for somatic gene therapy in mammals including humans. Furthermore, they are

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Principle for the Construction of Procon Vectors for targeted gene expression

Further, the replacement of β -galactosidase with a therapeutic gene such as one encoding melittin, cecropin or another antimicrobial peptide will result in a retroviral vector that can be manipulated to express this gene from any inserted promoter.

Procon v ctors carrying tissues specific promoters and regulatory elements such as the Tat Responsive Element

(TAR) from HIV will be useful for directing the expression of the therapeutic naturally occurring antimicrobial peptide sequences or derivatives thereof to predefined cell types, tissues and organs. Potential therapeutic sequences include mellitin, which has anti-HIV and anti-tumour effects, cecropin and megainin sequences and sequences which prime cells for death including the thymidine kinase, guanine phosphoribosyltransferase and cytosine deaminase genes.

10 BRIEF DESCRIPTION OF THE FIGURES

Figure 1 is a schematic of the mode of reverse transcription of a retrovirus.

Figure 2 is a schematic of retroviral vector constructs carrying melittin and cecropin coding sequences.

15 Figure 3 is a schematic of the construction of U3
minus BAG-vector (MLV).

Figures 4A-4C are schematics of the construction of the retroviral vector p125.Cecr.A carrying the PreProCecropin A gene.

20 Figures 5A-5C are schematics of the construction of the retroviral vectors pBAGpMel and pBAGppMel carrying respectively the PreMelittin and the PreProMelittin gene.

Figure 6 is a schematic of the pBAGpMel and pBAGppMel.

Figure 7 is a schematic of the pBAG.

25 Figures 8A-8B are schematics of the principle of S1 analysis of transfected Cecropin and Melittin clones.

Figures 9A-9G are graphs of days after injection versus tumor site showing antitumour activity of retroviral vectors carrying PreProCecropin coding sequences.

30 Figures 10A-10F are graphs of days after injection
versus tumor site showing antitumour activity of retroviral
vectors carrying Melittin coding sequences.

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BAG vector has been constructed in which the MLV promoter (U3) located within the 3'LTR (Figure 3) has been deleted by PCR. At this position a polylinker was inserted containing the restriction sites *Sac*II and *Mlu*I allowing the facile introduction of heterologous promoters. The BAG vector lacking the U3 is expressed from the MLV promoter (U3) within the 5'LTR when introduced into a packaging cell line. As a result of the rearrangements occurring in the retroviral genome during its life cycle, following infection of its target cell, the polylinker will be duplicated at both ends of the retroviral genome as described in Danish patent application no. 1017/94. Thereby a retroviral vector can be constructed in which the expression of the β -galactosidase gene of BAG will be controlled by the polylinker or any promoter inserted into the polylinker in the target cell (Figure 3).

The Mouse Mammary Tumour Virus (MMTV) U3-region (mtv-2) without the inverted repeats, which contains the MMTV promoters as well as a region that confers responsiveness to glucocorticoid hormones and a region containing an element that directs expression to the mammary gland was inserted into the polylinker region of the modified BAG vector to produce p125gal (Figure 4A).

Construction of p125CecrA

Figures 4A-4C illustrate the construction of the retroviral vector p125CecrA. The plasmid pCecrA (obtained from Boman, H.G. et al., *Ann. Rev. Immunol.*, 13:1-51 (1995)) and illustrated in Figure 4A carries the cDNA of the PreProCecropin A gene from the giant silk moth (*Hyaophora cecropia*) cloned in front of the bacterial *sp6* promoter. The PreProCecropin sequence was obtained by

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amplification by PCR from this plasmid using the following primers:

Primer 1: CecrA1 5'-TATGACGTC-TCGTTAGAACGCGGCT-3'
(SEQ ID NO: 3)

5 Primer 2: CecrA3 5'-GGCAGATCT-TAAATGTATCATGCAAT-3'
(SEQ ID NO: 4)

CecrA1 carries an AatII restriction site (GACGTC) (SEQ ID NO: 5) in the 5' extension with a 3 base pair protection (TAT). Similarly CecrA3 carries a BglII site (AGATCT) (SEQ ID NO: 6) in the 5' extension with a 3 base pair protection (GGC).

The PCR product (370 bps) was then digested with AatII and BglII to make the restriction sites available for cloning (Figure 4B).

15 P125gal was digested with the restriction enzymes AatII and BamHI resulting in a fragment of 5043 base pairs (Figure 4B) and ligated to the fragment carrying the PreProCecropin A gene from pCerca. This resulted in the formation of p125.Cerca (Figure 4C), the BamHI/BglII sites
20 being lost.

Construction of retroviral vectors carrying the PrePreMelittin and PreMelittin sequence

Figures 5A-5C illustrate the construction of the retroviral vectors pBAGpMel and pBAGppMel. The plasmid
25 pUM13/4 (Vlasak, R., Uger-Ullmann, C., Keil, G. and Frischauf, A-M, *Eur. J. Biochem.*, 135:123-126 (1989)) (Figure 5A) carries the cDNA encoding the PrePromelittin gene from *apis mellifera*. The PreProMelittin and the Premelittin sequence was obtained by amplification by PCR
30 using the following primers:

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- Primer 1: 5'-ATAGACGTC-AAGGAAGGAAGCGATCGGA-3' (SEQ ID NO: 7)
- Primer 2: 5'-TATGGATCC-AACCCTGTTGCCTCTTACG-3' (SEQ ID NO: 8)
- 5 Primer 3: 5'-TCTTACATCTATGCG-GGAATTGGAGCAGTTCTGAA-3' (SEQ ID NO: 9)
- Primer 3': 5'-AACTGCTCCAATTCC-CGCATAGATGTAAGAAATGT-3' (SEQ ID NO: 10)

Primer 1 carries an AatII site (GACGTC) (SEQ ID NO: 11) in the 5' extension with an ATA 3 bp protection and primer 2 carries BamHI site (GGATCC) (SEQ ID NO: 12) in the 5' extension with a TAT 3 bp protection.

A PCR on the plasmid pUM13/4 with primers 1 and 2 results in the amplification of the whole PreProMelittin sequence (Figure 5B) carrying both AatII and BamHI sites to enable cloning into the retroviral vector pBAG (Figure 5A) after digestion of the plasmid with AatII and BamHI (Figure 5C). The resulting plasmid pBAGppMel is shown in Figure 6.

The amplification of the PreMelittin sequence was carried out using a combination of conventional PCR and recombinant PCR. Primer 3, though binding in the melittin gene, carries a 5' extension that corresponds to the 3' sequence of the Pre sequence. Similarly, the Primer 3', though binding in the 3' region of the Pre sequence, carries a 5' extension corresponding to the sequence in the 5' region of the melittin gene. Initially a PCR was made with either primer pair 1 and 3' or 2 and 3. The products from these two reactions were then used to make a recombinant PCR. This entailed hybridising the two PCR products with each other using the 5' extensions that were carried in the primers 3 and 3'. Subsequent addition of primers 1 and 2, which adds the AatII and BamHI sites for cloning into pBAG allowed the amplification of the entire

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PreMelittin sequence (Figure 5B). Plasmid pBAGpMel (Figure 6) was produced by digestion of this PCR product and pBAG with AatII and BamHI followed by ligation of the fragments.

Isolation of clones

- 5 EJ cells were transfected with p125.CercaA and the clones expressing neomycin resistance were isolated. Six clones were isolated: Cecropin A1.4, Cecropin A1.7, Cecropin A1.8, Cecropin A10.3, Cecropin A10.4, and Cecropin A10.8.
- 10 Similarly Ej cells were transfected with pBAGpMel and pBagpppMel and the clones expressing neomycin resistance were isolated. Three clones containing the premelittin gene were isolated: Pre Melittin 1, Pre Melittin 4, and Pre Melittin 6. Two clones containing prepromelittin were
- 15 isolated: Pre Pro Melittin 1, and Pre Pro Melittin 5.

S1 analysis of transfected Cecropin/Melittin clones

- An S1 analysis was performed using pBAG digested with SpeI as a probe (Figure 7), the resulting 6.1 kb fragment comprising 3'LTR, polyoma early region and 5'LTR. The
- 20 fragment was then end-labeled using polynucleotide kinase with 32P and hybridized against total RNA isolated from Ej (human bladder carcinoma cell line) cell clones that had been transfected with retroviral vector constructs bearing either preprocecropin (clones A1.4, A1.7, A1.8, A10.4, A10.3, and A10.8), premelittin (clones 1, 4 and 6), and
- 25 prepromelittin (clones 1 and 5) or the empty vector alone (pBAG clones 3 and 6) (Figure 8A).

- There followed an S1 nuclease digestion (Figure 8B), the enzyme cleaving only single stranded nucleic acids,
- 30 resulting in a 288 bp fragment arising from the hybridization of the probe to the mRNA coming from the

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Subsequent separation using a 6% polyacrylamide gel and exposure of the gel using a Phosphor-imaging system (Fuji BAS1000) revealed the expected bands.

Many amphipathic polypeptides are synthesized in a preproform which is inactive (Boman H.G., *Ann. rev. Immunol.*, 13:1-51 (1995)). The endopeptidase that cleaves off the presignal peptide in a co-translational process is thought to be present in all cells whereas the protease that converts promelittin to the active melittin form appears to be present only in certain cells (Kreil G., et al., *Eur. J. Biochem.*, 111:49-58 (1980)).

25 experiment the Cecropin A1.4 clone was tested in 4 mice,
only of which showed a tumour.

EJ derived cell clones carrying the melittin or cecropin expression vector or the parental BAG vector, not carrying a therapeutic gene, were supertransfected with an indicator construct carrying the HIV LTR (and thus the HIV

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promoter) linked to a firefly luciferase reporter gene (HIV-luc) in the absence or presence of a separate construct expressing Tat. In the absence of Tat there was little luciferase activity detectable from the HIV-luc
5 construct in all cell clones as expected since the HIV promoter requires Tat for its activity. In contrast, in the presence of Tat, cell clones carrying BAG or a premelittin carrying construct show significant levels of luciferase expression, whereas a cell clone transfected
10 with either a prepromelittin or a cecropin expression construct showed little luciferase expression (Figures 11 and 12). This suggests that cecropin, prepromelittin and to a lesser extent premelittin inhibit the Tat driven expression from the HIV LTR. Thus the production of HIV
15 from infected cells will be inhibited by the antimicrobial peptide gene carrying therapeutic retroviral vector. This effect is expected to lead to a lack of virus production from HIV infected cells.

A similar experiment to that of Figure 11 and 12 can
20 be seen in Figures 13A-13B whereby two different MMTV LTR's controlling the expression of a luciferase reporter gene (Wintersberger et al., *Proc. Natl. Acad. Sci. USA*, 92: 2745-2749) were separately supertransfected into cells carrying either PreProCecropin or PreProMelittin under
25 control of an MLV promoter. These experiments showed that the downregulatory effect of Cecropin is not restricted to the HIV promoter, but also work with another retroviral vector.

In conclusion the present invention provides
30 therapeutic products for the treatment of retroviral infections including HIV, tumours, bacterial and viral infections comprising vector constructs carrying genes or

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derivatives thereof of therapeutic active peptides including those for melittins, cecropins and magainins.

EQUIVALENTS

While this invention has been particularly shown and described with references to preferred embodiments thereof, it will be understood by those skilled in the art that various changes in form and details may be made therein without departing from the spirit and scope of the invention as defined by the appended claims. Those skilled in the art will recognize or be able to ascertain using no more than routine experimentation, many equivalents to the specific embodiments of the invention described specifically herein. Such equivalents are intended to be encompassed in the scope of the claims.

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